

Original Research



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Safe and Efficient Preparative Approach to Chiral α-Chloroketones Based on In-Flow Generated Diazomethane

Abstract

 α -Chloroketones are valuable semi-products in the organic synthesis and pharmaceutical industry. In particular, they are used as key building blocks for the production of HIV protease inhibitors, such as atazanavir and darunavir. A well-known approach to their synthesis involves the Arndt-Eistert homologation, which relies on the formation of diazoketones followed by their halogenation. However, this process poses significant safety and implementation challenges due to the use of diazomethane (CH₂N₂). The high toxicity, carcinogenicity, and explosion hazard of CH₂N₂ limit its large-scale application and require design of specialized laboratory setups to reduce the risks.

In this study, we present a new continuous-flow diazomethane generator that integrates the membrane technology with a traditional flow reactor setup for a safe and efficient generation of CH_2N_2 . The flow technology eliminates the need for storage and handling of diazomethane, while facilitating its direct use in multistep synthesis. As a proof-of-concept, we demonstrate its application in the three-step synthesis of chiral α -chloroketones from N-protected amino acids. The approach newly developed offers a safer, more efficient, and scalable alternative to conventional diazomethane-based processes, paving the way for broader industrial applications.

 $\textit{Keywords:}\$ flow processes; diazomethane; α -chloroketones; diazoketones; halomethylation; chiral compounds

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Безпечний та ефективний препаративний підхід до хіральних α -хлорокетонів на основі діазометану, що синтезується в проточному режимі

Анотація

 α -Хлорокетони є цінними напівпродуктами в органічному синтезі та у фармацевтичній промисловості. Зокрема, їх використовують як ключові будівельні блоки для отримання інгібіторів ВІЛ протеази, як-от атазанавір і дарунавір. Широко відомий підхід до їх синтезу передбачає гомологізацію Арндта-Айстерта, яка базується на утворенні діазокетонів із подальшим галогенуванням. Однак цей процес створює значні проблеми з безпекою та впровадженням через використання діазометану ($\mathrm{CH_2N_2}$). Висока токсичність, канцерогенність і вибухонебезпечність $\mathrm{CH_2N_2}$ обмежують його широкомасштабне застосування та вимагають створення спеціального лабораторного устатковання для зниження ризиків.

У цьому дослідженні ми репрезентуємо новий проточний генератор діазометану, який об'єднує мембранні технології з класичним реактором проточного синтезу. Ця система забезпечує безпечне та ефективне генерування CH_2N_2 , усуваючи потребу у його зберіганні та транспортуванні й одночасно полегшуючи його пряме використання в багатоетапному синтезі. Як доказ концепції ми демонструємо застосування реактора в тристадійному синтезі хіральних α -хлорокетонів із N-захищених амінокислот. Нещодавно розроблений підхід пропонує безпечнішу, ефективнішу та придатну до масштабування альтернативу звичайним процесам на основі діазометану, прокладаючи шлях до його більш широкого промислового застосування.

Ключові слова: потокові процеси; діазометан; α-хлорокетони; діазокетони; галогенування; хіральні сполуки

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Supporting information: Copies of ¹H and ¹³C NMR spectra.

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■ Introduction

Over the past two decades, a series of highly potent, orally bioavailable HIV protease inhibitors have been developed and approved for clinical use [1, 2]. These inhibitors, including atazanavir and darunavir, play a crucial role in highly active antiretroviral therapy (HAART) and are listed by the World Health Organization as essential medicines [3]. Structurally, they belong to the class of peptidomimetics, mimicking natural substrates by incorporating non-hydrolyzable hydroxyethylene or hydroxyethylamine moieties at the protease cleavage site (**Figure 1**). Notably, most FDA-approved HIV protease inhibitors contain a chiral amino alcohol core, which is typically introduced through a nucleophilic ringopening of the corresponding N-protected aminoepoxide [4].

A well-established route for preparing these chiral semi-products involves the transformation

of N-protected amino acids into α -haloketones, followed by a selective reduction to produce chiral amino epoxides or amino alcohols. Among various methods available for the α-haloketone formation, one of the most straightforward and costeffective strategies relies on the halomethylation using diazomethane (CH₂N₂) (**Scheme 1**) [5]. This method, involving the condensation of an activated amino acid with CH2N2 followed by the α,α-substitution with a hydrogen halide, is known due to its efficiency and diastereoselectivity [6]. However, diazomethane, an extremely versatile reagent in organic synthesis, is also highly toxic, volatile, carcinogenic, and prone to explosive decomposition, which pose significant limitations on its large-scale implementation. Its sensitivity to heat, light, and mechanical shock necessitates careful handling, and even the use of specialized equipment does not guarantee the avoidance of accidents [7]. Therefore, the industrial-scale production of CH₂N₂ could be provided using batch

Figure 1. HIV protease inhibitors derived from α -haloketones

Scheme 1. The diazomethane-based synthesis of α -haloketones from N-protected chiral amino acids

and continuous flow processes, with notable examples including the Aerojet process and other industrial setups utilizing the phase-transfer catalysis or continuous nitrogen stream transport [8]. Despite these advancements, the large-scale application of diazomethane remains restricted due to the challenges of safe storage, transport, and handling. To overcome these limitations, recent developments in flow chemistry have introduced safer, on-demand CH₂N₂ generation methods, integrating the membrane separation technology to minimize the risks associated with its accumulation [9, 10]. Continuous flow systems have long been employed for the large-scale chemical production and are being increasingly adapted for fine chemical and pharmaceutical syntheses [11]. These systems enable controlled multistep transformations, while ensuring greater safety, reproducibility, and efficiency.

In this study, we present the development of a procedure based on a continuous-flow generated diazomethane, applied to the three-step synthesis of α -chloroketones from N-protected amino acids. This approach eliminates the need for diazomethane storage and handling while enabling a scalable and efficient synthesis of some key chiral building blocks without racemization. The full continuous process demonstrates high yields and safety improvements, making it a viable alternative to traditional batch methods for producing α -haloketones in pharmaceutical applications.

Results and discussion

Generation of Anhydrous Diazomethane

Access to anhydrous diazomethane (CH_2N_2) is essential for the modified Arndt-Eistert reaction, which serves as a key step in the synthesis of chiral amino alcohols – crucial semi-products in the production of HIV protease inhibitors. Ensuring the complete removal of water from CH_2N_2 is critical as even trace moisture can affect the reaction efficiency and selectivity.

Previous studies have employed the reaction between $\mathrm{CH_2N_2}$ and activated organic acids as an indirect method to verify anhydrous conditions [12]. However, our investigations revealed that this technique lacks the required sensitivity.

When benzoyl chloride was exposed to $\mathrm{CH_2N_2}$ under controlled conditions, the expected formation of benzoic acid (a marker for water presence) was only detectable by GC-FID when at least 50 vol% of water was added.

To obtain more accurate moisture measurements, we turned to the Karl-Fischer titration, a highly sensitive technique for water quantification [12]. In our setup, a 0.1 M CH₂N₂ solution in the THF:CH₂Cl₂ mixture was generated and subsequently quenched with benzoic acid. A reference Karl-Fischer titration was performed on a pure benzoic acid solution, and this baseline value was subtracted from the reading obtained for the quenched CH₂N₂ solution. The final titration results confirmed that our system successfully produces anhydrous CH2N2, with a measured water content of 347 ± 7 ppm, demonstrating that the membrane-based diazomethane generation effectively prevents the moisture contamination.

By ensuring a continuous supply of high-purity $\mathrm{CH_2N_2}$, our method provides a safe, efficient, and scalable alternative to conventional batch approaches, further enhancing its applicability in multistep synthetic processes.

Synthesis of a-Chloroketones

The synthesis of α -chloroketones was carried out via a well-established three-step pathway comprising the activation of the N-protected amino acid, the formation of the diazoketone semi-product, and the selective halogenation to yield the final product. This methodology, originally optimized for the α -bromoketone synthesis in the previous study [10], was successfully adapted for the α -chloroketone formation by substituting hydrogen bromide (HBr) with hydrogen chloride (HCl) as a halogenating agent.

Step 1: The Activation of N-Protected Amino Acids

In the first step, *N*-protected amino acids 1 were converted into reactive mixed anhydrides 1-act by the treatment with ethyl chloroformate in the presence of a base, such as triethylamine or *N*-methylmorpholine, in an anhydrous solvent (**Figure 2**, *A*). The procedure was performed under mild reaction conditions that avoided excessive heating or drastic pH changes (*see* Experimental section), and it was compatible with

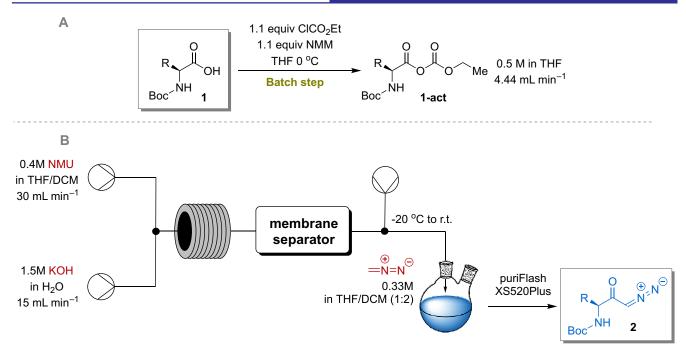


Figure 2. The intermediate synthesis (A) and reactor setup (B)

a broad range of amino acid protecting groups, thereby offering flexibility in the precursor selection. Moreover, the reaction proceeded with a high conversion efficiency, which minimized side reactions and the by-product formation.

Step 2: The Formation of the a-Diazoketone

Once the mixed anhydride was generated, it reacted with anhydrous diazomethane (CH₂N₂) under carefully controlled conditions. By employing a continuous-flow CH_2N_2 generator (**Figure 2**, B), a high-purity stream of diazomethane was directed into the reaction mixture, ensuring a rapid and efficient conversion of the mixed anhydride to the corresponding α-diazoketone. The continuousflow approach enhanced safety by eliminating the need for accumulation and storage of a large amount of diazomethane, provided a high reproducibility through the consistent conversion efficiency across various substrates, and improved the reaction kinetics by facilitating a rapid formation of the diazoketone. The formation of the diazoketone intermediates was confirmed via ¹H and ¹³C NMR spectroscopy, with characteristic peaks in agreement with literature reports for similar compounds [9, 10].

Step 3: The Halogenation to a-Chloroketones In the final step, α-diazoketone semi-products were selectively converted into the corresponding α-chloroketones using aqueous HCl. Chlorination, unlike bromination where rapid elimination side reactions can occur, benefited from the lower nucleophilicity of chloride ions, which enhanced selectivity. Initial attempts focused on developing a fully continuous α-chloroketone synthesis

using a flow reactor. However, challenges emerged since the high surface tension of the aqueous HCl phase impeded a reliable flow control, which in turn led to uncontrolled increases in solvent levels. Drawing on insights from previous work, the process was modified by transitioning to a conventional batch synthesis approach. In this modified protocol, a pure isolated diazoketone obtained via the continuous-flow process in Step 2 was used as the starting material for the halogenation. The isolated diazoketone reacted with 3 equiv. of the conc. HCl at room temperature. The complete conversion was achieved within 10 minutes of stirring. It is important to note that increasing the HCl quantity beyond 3 equiv. resulted in the unwanted deprotection of α -chloroketone, underscoring the necessity for the precise reagent control. The final α-chloroketones were isolated with purities exceeding 98%, as confirmed by the HPLC analysis.

The chloroketones prepared by this protocol are shown in **Figure 3** (*B*). The protocol was easily scaled up to 100 g of chloroketone in a single synthetic run. We developed individual protocols for the optical purity control based on the chiral HPLC for all chiral representatives. As a result, we found that the optical purity of the starting amino acids was retained for all chloroketones. It also proves the preservation of the absolute configuration for the intermediate diazoketones.

Comparison with Alternative Methods

The performance of the elaborated method was evaluated against other reported approaches based on the diazoketone formation and

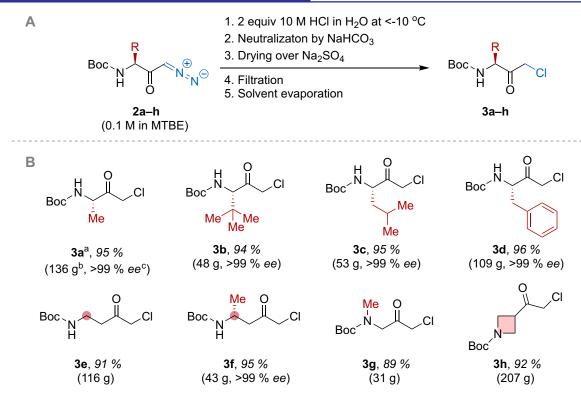


Figure 3. The general scheme for the preparation of chloroketones from diazoketones (*A*); the scope of chloroketones **3a-h** (*B*) prepared. ^a For all chiral products, both enantiomers were independently synthesized (*see* **Experimental section**). ^b The amount of the compound obtained. ^c Measured using chiral HPLC.

halogenation time, yield, purity, and overall safety profile. The classic batch method, which employs pre-made CH₂N₂, typically requires 2–3 hours for the halogenation, with yields of products in the range of 65–75%, the purity between 90–95% and is associated with a high safety risk. The tube-intube method described by Pinho et al. [13] uses a continuous diazoketone formation and achieves the halogenation in 1-2 hours, with yields of 75–85% and the purity of 95–98%, presenting a moderate safety profile. In contrast, our method, which combines a continuous CH₂N₂ generation in Step 2 with the batch halogenation in Step 3, achieves the diazoketone formation within 30-60 minutes and the halogenation in 10 minutes, resulting in yields of 85-92% and the purity higher than 98% while offering a high safety profile.

Conclusion

By adapting our bromoketone synthesis method previously reported for the chloroketone preparation, we have developed a preparative process that combines the continuous-flow generation of α -diazoketones with the subsequent batch halogenation step. This hybrid approach enhances scalability and reproducibility while maintaining a high safety profile by eliminating the need for the hazardous diazomethane accumulation,

storage and handling. Taking into account the significant role of α -haloketones in pharmaceutical applications, our methodology represents a noteworthy advance in flow chemistry and a new synthetic employment of easily available diazo compounds.

Experimental section

General

The solvents were purified according to standard procedures. All starting materials were obtained from Enamine Ltd. ¹H and ¹³C NMR spectra were recorded on a Bruker Avance 500 spectrometer (500 MHz for ¹H and 126 MHz for ¹³C nuclei) and a Varian Unity Plus 400 spectrometer (400 MHz for ¹H and 101 MHz for ¹³C nuclei). Tetramethylsilane was used as an internal standard. Melting points were measured on the MPA100 OptiMelt automated melting point system. HPLC analyses were performed on an Agilent 1200 chromatograph. A preparative chromatograph puriFlash XS520Plus with a column containing 800 g of silica gel was used for diazoketones purification.

General Methods for the Synthesis of Compounds

The **NMU** precursor was prepared by nitrosation of methylurea using the standard protocol [14].

The Synthesis of Mixed Anhydrides

In a three-necked flask equipped with a thermometer, a magnetic stirrer, a dropping funnel, and a water seal, 1 equiv. of methylmorpholine was added. The reaction mixture was cooled below -10 °C and then 1 equiv. of ethyl chloroformate was slowly added, keeping the temperature in the flask below 0 °C. The reaction mixture was stirred at this temperature for 30 min. The precipitate formed was filtered off, and the solution of the mixed anhydride was used in the further transformations without isolation.

The Synthesis of Diazoketones

The procedure of diazomethane generation Solutions of NMU (0.37 M, $CH_2Cl_2/THF = 2:1$) and KOH (1.5 M, H₂O) were injected into the reaction unit through two channels of the reactor pump unit in a molar ratio of 1:2 (30 and 15 mL min⁻¹). The combined stream passed through the reaction column and then through the membrane separator. The aqueous waste stream leaving the outer tube of the separator was directed into a flask containing acetic acid to decompose any residue of diazomethane in the aqueous solution. The resulting organic solution of diazomethane was transferred from the inner tube to a reactor containing a substrate. The flux of diazomethane (0.45 mol h⁻¹) was directed into a three-necked flask equipped with a magnetic stirrer and an electronic thermometer. The substrate flow kept in a molar ratio of 1:2 in relation to diazomethane was fed into the reactor with a delay of 1 min. After the entire amount of the substrate was added, the reactor was switched to wash the mode, and the reaction mixture was stirred for an additional 2 h. The system was able to operate continuously for 5–6 h (stationary conditions) generating and consuming up to 1.8 mol of diazomethane. After evaporation of solvents under vacuum, diazoketone was purified using a preparative chromatograph puriFlash XS520Plus and hexane with a gradient addition of ethyl acetate up to 30% as an eluent. The total amount of diazoketone after the isolation and purification exceeded 1 mol.

The synthesis of chloroketones

Diazoketone was dissolved in MTBE in a threenecked flask equipped with a thermometer, a magnetic stirrer, a dropping funnel and a water seal. The solution was cooled with ice to a temperature below 10 °C. Two (2) equiv. of the concentrated HCl was slowly added, keeping the temperature in the flask below 10 °C. The reaction mixture was stirred at this temperature for 45 min. The resulting solution was neutralized with aq. NaHCO₃ to pH = 7-8, washed with water, dried over Na₂SO₄ and filtered. The solvent was evaporated under reduced pressure. The purity of the chloroketones prepared was >95%, no further purification was necessary.

(S)-tert-Butyl (4-Chloro-3-oxobutan-2-yl)-carbamate (3a)

A white powder. Yield -136 g (95%). M. p. = 59–60 °C. Anal. Calcd for $C_9H_{16}ClNO_3$, %: C 46.85, H 6.94, N 5.26, Cl 15.40. Found, %: C 46.63, H 6.93, N 5.12, Cl 15.07. ¹H NMR (400 MHz, CDCl₃), δ , ppm: 1.41 (d, J = 7.2 Hz, 3H), 1.46 (s, 9H), 4.08 (d, J = 4.0 Hz, 2H), 4.50–4.67 (m, 1H), 5.10 (s, 1H). ¹³C NMR (151 MHz, CDCl₃), δ , ppm: 17.77, 28.28, 31.61, 53.22, 80.30, 155.16, 201.53. [α]_D(25°C) = -54.2 (c = 0.5, CH₃OH).

(R)-tert-Butyl (4-Chloro-3-oxobutan-2-yl)-carbamate (3a')

A white powder. Yield -37 g (94%). M. p. = 59-60 °C. [α]_D = +54.2 (c = 0.5, CH₃OH).

(S)-tert-Butyl (1-Chloro-4,4-dimethyl-2-oxopentan-3-yl)-carbamate (3b)

A white crystalline powder. Yield – 48 g (94%). M. p. = 82 °C. Anal. Calcd for $C_{12}H_{22}ClNO_3$, %: C 52.84, H 8.07, N 5.14, Cl 13.03. Found, %: C 52.51, H 7.88, N 4.92, Cl 13.13. ¹H NMR (400 MHz, CDCl₃), δ , ppm: 0.98 (s, 9H), 1.41 (s, 9H), 4.05–4.23 (m, 2H), 4.30 (d, J = 9.0 Hz, 1H), 5.07 (d, J = 9.1 Hz, 1H). ¹³C NMR (151 MHz, CDCl₃), δ , ppm: 26.5, 28.3, 34.6, 36.2, 64.0, 80.3, 155.5, 201.5. [α]_D(25°C) = -47.0 (c = 0.5, CH₃OH).

(*R*)-*tert*-Butyl (1-Chloro-4,4-dimethyl-2-oxopentan-3-yl)-carbamate (3b')

A white powder. Yield -25 g (94%). M. p. = 82 °C. $[\alpha]_D(25 \text{ °C}) = +48.1 \text{ (c} = 0.5, \text{ CH}_3\text{OH})$.

(*R*)-*tert*-Butyl (1-Chloro-5-methyl-2-oxohe-xan-3-yl)-carbamate (3c)

A white powder. Yield -53 g (95%). M. p. = 76 °C. Anal. Calcd for $C_{12}H_{22}ClNO_3$, %: C 52.84, H 8.07, N 5.14, Cl 13.03. Found, %: C 52.53, H 7.89, N 4.95, Cl 13.17. 1H NMR (400 MHz, CDCl₃), δ , ppm: 0.98 (t, J=6.7 Hz, 6H), 1.46 (s, 10H), 1.56–1.66 (m, 1H), 1.68–1.84 (m, 1H), 3.98–4.24 (m, 2H), 4.55 (s, 1H), 4.92 (d, J=8.0 Hz, 1H). ^{13}C NMR (126 MHz, CDCl₃), δ , ppm: 21.1, 22.7, 24.5, 27.8, 31.8, 40.2, 55.6, 79.9, 155.0, 201.3. [α]_D(25°C) = -45.4 (c = 1.0, CH₃OH).

(R)-tert-Butyl (1-Chloro-5-methyl-2-ox-ohexan-3-yl)-carbamate (3c')

A white powder. Yield – 23 g (95%). [a] $_{D}(25^{\circ}\text{C}) = +44.5 \text{ (c} = 1.0, \text{CH}_{3}\text{OH}).$

(S)-tert-Butyl (4-Chloro-3-oxo-1-phenyl-butan-2-yl)-carbamate (3d)

A white powder. Yield – 109 g (96%). M. p. = 101 °C. Anal. Calcd for $C_{15}H_{20}ClNO_3$, %: C 58.73, H 6.52, N 4.57, Cl 11.58. Found, %: C 58.63, H 6.50, N 4.55, Cl 11.38. ¹H NMR (400 MHz, CDCl₃), δ , ppm: 1.42 (s, 9H), 3.08 (tt, J=20.7, 10.0 Hz, 2H), 3.79–4.13 (m, 2H), 4.73 (q, J=6.5, 6.1 Hz, 1H), 5.04 (s, 1H), 7.13–7.23 (m, 2H), 7.27 (s, 1H), 7.33 (q, J=9.0, 7.9 Hz, 2H). ¹³C NMR (151 MHz, CDCl₃), δ , ppm: 28.2, 33.2, 37.9, 58.5, 80.5, 127.3, 128.9, 129.1, 135.8, 155.2, 200.8. [α]_D(25°C) = -43.7 (c = 0.5, CH₃OH).

(R)-tert-Butyl (4-Chloro-3-oxo-1-phenyl-butan-2-yl)-carbamate (3d')

A white powder. Yield – 29 g (97%). $[\alpha]_D$ (25°C) = +55.4 (c = 0.5, CH₃OH).

tert-Butyl (4-Chloro-3-oxobutyl)carbamate (3e)

A colorless liquid. Yield – 116 g (91%). Anal. Calcd for C₉H₁₆ClNO₃, %: C 46.85, H 6.94, N 6.07, Cl 15.40. Found, %: C 46.71, H 6.90, N 6.17, Cl 15.37. ¹H NMR (400 MHz, CDCl₃), δ , ppm: 1.44 (s, 9H), 2.91 (t, J=5.8 Hz, 2H), 3.37–3.51 (m, 2H), 3.91 (s, 2H), 4.96 (s, 1H). ¹³C NMR (126 MHz, CDCl₃), δ , ppm: 27.9, 33.6, 34.9, 39.6, 79.0, 155.4, 201.0.

(S)-tert-Butyl (5-Chloro-4-oxopentan-2-yl)carbamate (3f)

A yellow powder. Yield -43 g (95%). M. p. = 73 °C. Anal. Calcd for $C_{10}H_{18}ClNO_3$, %: C 49.08, H 7.36, N 5.73, Cl 14.52. Found, %: C 49.18,

H 7.27, N 5.77, Cl 14.12. ¹H NMR (400 MHz, CDCl₃), δ, ppm: 1.23 (d, J = 7.0 Hz, 3H), 1.43 (s, 9H), 2.86 (d, J = 5.8 Hz, 2H), 3.92 (q, J = 12.6 Hz, 2H), 4.02–4.15 (m, 1H), 4.78 (s, 1H). ¹³C NMR (126 MHz, CDCl₃), δ, ppm: 20.1, 27.9, 34.0, 43.0, 45.7, 79.1, 154.6, 200.1. [α]_D(25°C) = −16.8 (c = 0.5, CH₃OH).

(R)-tert-Butyl (5-Chloro-4-oxopentan-2-yl)carbamate (3f')

A yellow powder. Yield -200 g (95%). M. p. = 73 °C. $[\alpha]_D(25\text{°C}) = +16.1 \text{ (c} = 0.5, \text{CH}_3\text{OH})$.

tert-Butyl (3-Chloro-2-oxopropyl)(methyl)carbamate (3g)

An orange oil. Yield – 31 g (89%). Anal Calcd for $C_9H_{16}ClNO_3$, %: C 46.85, H 6.94, N 6.07, Cl 15.40. Found, %: C 46.74, H 7.02, N 6.17, Cl 15.37. ¹H NMR (400 MHz, CDCl₃), δ , ppm: 1.46 (d, J=18.8 Hz, 9H), 2.94 (d, J=7.7 Hz, 3H), 3.91 (d, J=16.9 Hz, 2H), 4.22 (d, J=18.2 Hz, 2H). ¹³C NMR (151 MHz, CDCl₃), δ , ppm: 24.1*, 27.0, 28.2*, 28.3, 30.7*, 31.5, 35.9*, 36.4, 55.9*, 56.3, 80.5*, 80.7, 156.1, 198.4 (rotamers).

tert-Butyl 3-(2-Chloroacetyl)azetidine-1-carboxylate (3h)

A white crystalline powder. Yield -207 g (92%). M. p. = 73 °C. Anal. Calcd for $C_{10}H_{16}ClNO_3$, %: C 49.48, H 6.60, N 5.77, Cl 14.64. Found, %: C 49.45, H 6.50, N 5.72. Cl 14.63. 1H NMR (400 MHz, CDCl₃), δ , ppm: 1.44 (s, 9H), 3.82 (pd, J = 7.5, 1.2 Hz, 1H), 3.91 (s, 2H), 4.08 (s, 2H), 4.10 (s, 2H). ^{13}C NMR (126 MHz, CDCl₃), δ , ppm: 28.3, 32.5, 36.6, 51.1, 80.0, 156.0, 200.2.

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